

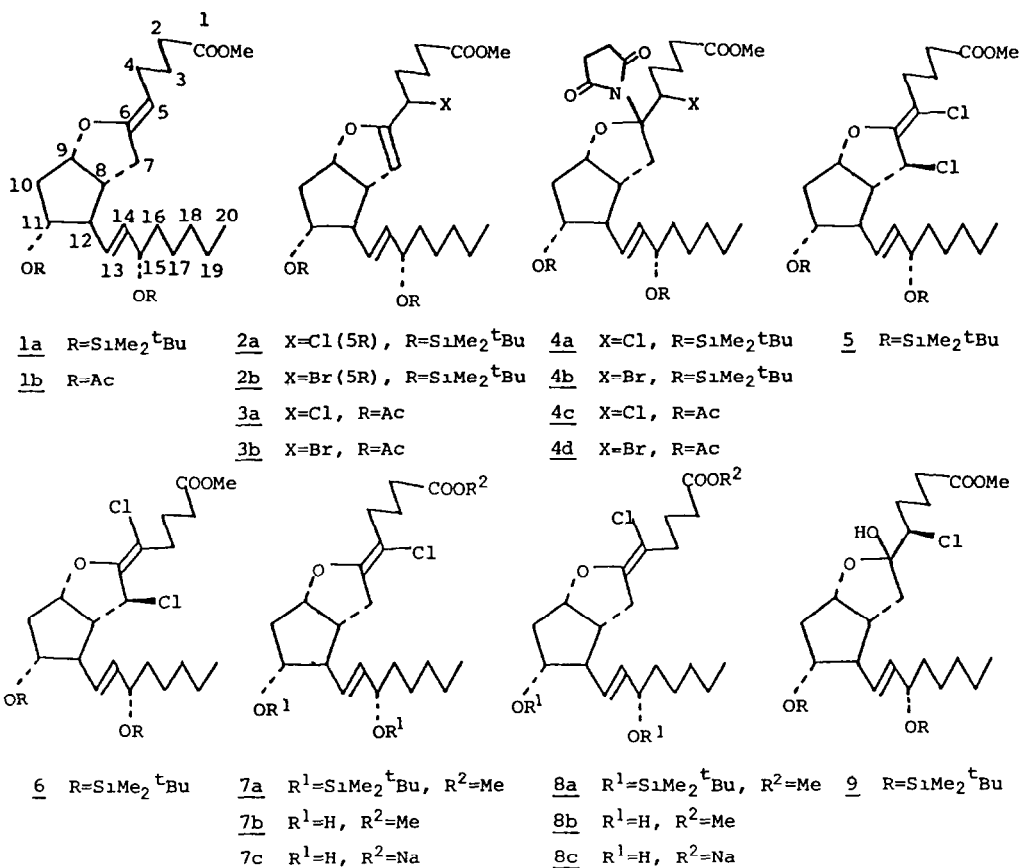
HALOGENATION OF PGI<sub>2</sub>-ENOL ETHER WITH N-HALOSUCCINIMIDE SYNTHESIS  
OF NEW STABLE PGI<sub>2</sub> ANALOGS, 5-CHLORO- AND 5,7-DICHLORO-PGI<sub>2</sub><sup>†</sup>

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*Reaction of the enol ether of PGI<sub>2</sub> with N-halosuccinimide and subsequent isomerization or halogenation gave new stable PGI<sub>2</sub> analogs, 5-chloro- and 5,7-dichloro-PGI<sub>2</sub>*

Many efforts have been made to obtain stable PGI<sub>2</sub> analogs which would be expected to be promising therapeutic agents<sup>1</sup>. One of the efficient methods to stabilize PGI<sub>2</sub> is to introduce (an) electron-withdrawing group(s) around the enol ether function in the molecule<sup>2</sup>. In this respect, we reported the syntheses of 5-PhS-,<sup>3</sup> 7-OH and 7-OAC-PGI<sub>2</sub>,<sup>4</sup> which were stable, however, unfortunately with weak biological activities. This time we chose (a) halogen atom(s) as (an) electron-withdrawing group(s), which would be introduced in the molecule of PGI<sub>2</sub> by the electrophilic halogenation. It is well documented that the electrophilic halogenation of silyl enol ethers with N-halosuccinimide gives  $\alpha$ -haloketones,<sup>5</sup> whereas the reaction of alkyl enol ethers has not been much studied.<sup>6</sup> We report here that the reaction of PGI<sub>2</sub> with N-halosuccinimide and subsequent isomerization or halogenation led to new stable PGI<sub>2</sub> analogs of 5-chloro- and 5,7-dichloro-PGI<sub>2</sub>.

Halogenation of *t*-butyldimethylsilyl-protected PGI<sub>2</sub> methyl ester 1a<sup>7</sup> with N-chloro- or N-bromosuccinimide (1.05 equiv, CCl<sub>4</sub>, r.t., 1 h) proceeded stereoselectively to afford silyl-protected (5*R*)-5-chloro- $\Delta^6$ -PGI<sub>1</sub> methyl ester 2a<sup>8</sup> or (5*R*)-5-bromo- $\Delta^6$ -PGI<sub>1</sub> methyl ester 2b in 60% or 57% yields accompanied with the formation of succinimido-adduct 4a (20%) or 4b (20%). Both compounds 2a and 2b were found homogeneous by their <sup>13</sup>C-NMR spectra and their 5*R*-configuration was tentatively assigned on the basis of the mechanistic consideration (*vide infra*). On the other hand, the similar halogenations of acetyl-protected PGI<sub>2</sub> methyl ester 1b gave a diastereomeric mixture of 5-chloro- $\Delta^6$ -PGI<sub>1</sub> methyl ester 3a (58%) (5*R* 5*S* = *ca* 5:3)<sup>9</sup> or 5-bromo- $\Delta^6$ -PGI<sub>1</sub> methyl ester 3b (60%) (5*R* 5*S* = *ca* 7:3)<sup>9</sup> besides respective succinimido-adduct 4c (20%) or 4d (13%). The selective formation of the 5*R*-isomers from protected PGI<sub>2</sub> methyl ester 1 could be accounted for the bulkiness of the C-11 substituent, which is close to the enol ether double bond as depicted in Fig. 1,<sup>10</sup> where the halogenating agent could approach the enol ether double bond more easily from its less hindered upper side than from the lower side resulting in the selective formation of 5*R*-isomer. Further chlorination of silyl-protected-(5*R*)-allyl chloride 2a was accomplished by treatment with N-chlorosuccinimide (10 equiv, CCl<sub>4</sub>, 60°C, 3 h)



to afford a geometric mixture of (5*E*,7*S*)- and (5*Z*,7*S*)-5,7-dichloro PGI<sub>2</sub> methyl esters 5 and 6 in 15% and 20% yields, respectively<sup>11</sup>

In order to obtain 5-chloro-PGI<sub>2</sub> derivative 7, in the hope that 7 would be one of the promising stable PGI<sub>2</sub> analogs, isomerization of the endo-double bond ( $\Delta^6$ ) of above obtained 5-Cl- $\Delta^6$ -PGI<sub>1</sub> 2a to exo-double bond ( $\Delta^5$ ) was examined. Previously we achieved to isomerize 5-PhS- $\Delta^6$ -PGI<sub>1</sub> 10 stereospecifically into 5-PhS-PGI<sub>2</sub> 11 in excellent yield<sup>4</sup> by treatment with trace conc H<sub>2</sub>SO<sub>4</sub>. However, the similar treatment of 5-Cl- $\Delta^6$ -PGI<sub>1</sub> 2a with trace conc H<sub>2</sub>SO<sub>4</sub> gave isomerized 7a and 8a only in low yield. After several examinations, the isomerization of 2a was effected as follows. Treatment of a moistured benzene solution of allyl chloride 2a with a catalytic amount of pyridinium *p*-toluenesulfonate (r t, 16 h) gave hemiacetal 9. Subsequent dehydration of 9 with excess anhydrous MgSO<sub>4</sub> in refluxing benzene (15 h)<sup>12</sup> afforded

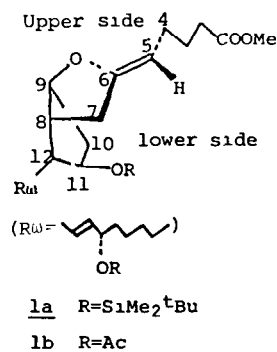


Fig. 1

5*E*-vinyl chloride 7a (20%) and 5*Z*-vinyl chloride 8a (28%) along with the starting allyl chloride 2a (18%). The silyl protecting groups of vinyl chlorides 7a and 8a were removed with *n*-Bu<sub>4</sub>NF-Et<sub>3</sub>N (15 equiv) in THF (r.t., 5 h) to afford 5-chloro-PGI<sub>2</sub> methyl ester 7b (95%) and (5*Z*)-5-chloro-PGI<sub>2</sub> methyl ester 8b (80%). Solutions of sodium salts 7c and 8c were prepared by hydrolysis of 7b and 8b with NaOH-EtOH-H<sub>2</sub>O (r.t., 3 h). The  $\Delta^5$ -olefin geometry of 5-chloro-PGI<sub>2</sub> methyl esters 7b and 8b, was determined on the basis of the <sup>1</sup>H-NMR (400 MHz) study with the aid of precise decoupling studies. The chemical shifts of the C-4 methylene protons of 7b were found to be 2.38 ppm (1H) and 2.48 ppm (1H), while those of 8b, 2.27 ppm (2H). Moreover, the C-7 methylene protons of 7b (7 $\alpha$ -H, 2.63 ppm, 7 $\beta$ -H, 2.70 ppm) appeared in a field lower than that of 8b (7 $\alpha$ -H, 2.52 ppm, 7 $\beta$ -H, 2.67 ppm). Considering the deshielding effect<sup>13</sup> of oxygen and chlorine atoms on *cis*-related allylic protons, the  $\Delta^5$ -olefin geometry of 7b and 8b were assigned to *E* and *Z*, respectively.

These PGI<sub>2</sub> analogs described here were found to be much more stable than PGI<sub>2</sub>. The half life of 7c and 8c in a pH 4.7 buffer solution were 1.5 h and 8 h, respectively,<sup>14</sup> while that of PGI<sub>2</sub> in a pH 5.98 buffer solution is only 22.4 seconds.<sup>15</sup> Analog 7c showed an inhibitory activity on platelet aggregation (IC<sub>50</sub>, 0.14  $\mu$ g/ml), while 8c was less active. The activity of 7c was retained after standing for 4 h at 0°C in a pH 7.4 buffer solution, whereas that of PGI<sub>2</sub> diminished to about 1/100 under the same condition.

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#### Spectral Data

2a <sup>1</sup>H-NMR(CDCl<sub>3</sub>)  $\delta$  2.85-3.2 (1H, br), 3.66(3H, s), 3.7-4.2(2H, m), 4.38(1H, t, *J*=7 Hz), 4.7-5.0 (1H, br), 5.00(1H, d, *J*=3 Hz), 5.52(2H, m), MS(70 eV) *m/e* 630, 628(M<sup>+</sup>), <sup>13</sup>C-NMR(CDCl<sub>3</sub>)  $\delta$  22.0(C-55), 55.4(C<sub>5</sub>), 76.1(C<sub>11</sub>), 83.3(C<sub>9</sub>), 101.6(C<sub>7</sub>), 154.8(C<sub>6</sub>). 3a MS(20 eV) *m/e* 486, 484 (M<sup>+</sup>), <sup>13</sup>C-NMR(CDCl<sub>3</sub>)  $\delta$  22.1 and 22.0(C<sub>3</sub>), 54.9(C<sub>5</sub>), 78.03 and 77.79(C<sub>11</sub>), 84.9 and 84.6(C<sub>9</sub>), 101.3 and 101.0(C<sub>7</sub>), 155.0 and 155.4(C<sub>6</sub>). 5 <sup>1</sup>H-NMR(CDCl<sub>3</sub>)  $\delta$  3.67(3H, s), 3.85(1H, q, *J*=8 Hz), 4.05-4.3 (1H, br), 4.79(1H, s), 4.8-5.1(1H, br), 5.53(2H, m), MS(70 eV) *m/e* 607, 605(M-57). 6 <sup>1</sup>H-NMR(CDCl<sub>3</sub>)  $\delta$  3.67(3H, s), 3.86(1H, q, *J*=7 Hz), 3.9-4.2(1H, br), 4.69(1H, s), 4.85-5.1(1H, br), 5.53(2H, m), MS(70 eV) *m/e* 607, 605(M-57). 7b <sup>1</sup>H-NMR(CDCl<sub>3</sub>)  $\delta$  2.16(1H, q, *J*=8 Hz, C<sub>12</sub>-H), 2.325(2H, t, *J*=7 Hz, C<sub>2</sub>-H<sub>2</sub>), 2.37(1H, q, *J*=8 Hz, C<sub>8</sub>-H), 2.38(1H, dt, *J*=14 Hz, 7 Hz, C<sub>4</sub>-H), 2.48 (1H, dt, *J*=14 Hz, 7 Hz, C<sub>4</sub>-H), 2.48(1H, dt, *J*=14 Hz, 7 Hz, C<sub>10</sub> $\alpha$ -H), 2.63(1H, d, *J*=17 Hz, C<sub>7</sub> $\alpha$ -H), 2.70(1H, dd, *J*=17 Hz, 8 Hz, C<sub>7</sub> $\beta$ -H), 3.68(3H, s), 3.90(1H, br, C<sub>11</sub>-H), 4.10(1H, q, *J*=7 Hz, C<sub>15</sub>-H), 4.66(1H, dt, *J*=3 Hz, 8 Hz, C<sub>9</sub>-H), 5.53(1H, dd, *J*=8 Hz, 15 Hz, C<sub>13</sub>-H), 5.64(1H, dd, *J*=6 Hz, 15 Hz, C<sub>14</sub>-H), MS(20 eV) *m/e* 402, 400(M<sup>+</sup>). 8b <sup>1</sup>H-NMR(CDCl<sub>3</sub>)  $\delta$  2.17(1H, q, *J*=8 Hz, C<sub>12</sub>-H), 2.27(2H, t, *J*=7 Hz, C<sub>4</sub>-H<sub>2</sub>), 2.33(2H, t, *J*=7 Hz, C<sub>2</sub>-H<sub>2</sub>), 2.44(1H, q, *J*=8 Hz, C<sub>8</sub>-H), 2.48 (1H, d, *J*=16 Hz, C<sub>7</sub> $\alpha$ -H), 2.53(1H, dt, *J*=14 Hz, 8 Hz, C<sub>10</sub>-H), 2.67(1H, dd, *J*=16 Hz, 8 Hz, C<sub>7</sub> $\beta$ -H), 3.67(3H, s), 3.91(1H, q, *J*=8 Hz, C<sub>11</sub>-H), 4.10(1H, q, *J*=7 Hz, C<sub>15</sub>-H), 4.74(1H, dt, *J*=3 Hz, 8 Hz, C<sub>9</sub>-H), 5.52(1H, dd, *J*=8 Hz, 16 Hz, C<sub>13</sub>-H), 5.64(1H, dd, *J*=6 Hz, 16 Hz, C<sub>14</sub>-H), MS(20 eV) *m/e* 402, 400(M<sup>+</sup>).

## References and Notes

- † Prostaglandin Chemistry XVIII For part XVII, K Bannai, T Toru, A Hazato, T Ōba, T Tanaka, N Okamura, K Watanabe, and S Kurozumī, *Chem Pharm Bull*, 30, 1102 (1982)
- 1 P J Lewis and J O'Grady (ed), "Clinical Pharmacology of Prostacyclin", Raven Press, New York, 1981
- 2 G Kovács, V Simonidez, I Tómoskozi, P Kormóczy, I Szekely, A Papp-Behr, I Stadler, L Szekeres, and Gy Papp, *J Med Chem*, 25, 105 (1982), J Fried, D K Mitra, M Nagarajan, and M M Mehrotra, *ibid*, 23, 234 (1980), Schering AG Ger Offen 2801846, Japan Kokai 54-81257
- 3 T Toru, K Watanabe, T Ōba, T Tanaka, N Okamura, K Bannai, and S Kurozumī, *Tetrahedron Lett*, 21, 2539 (1980)
- 4 K Bannai, T Toru, T Ōba, T Tanaka, N Okamura, K Watanabe, and S Kurozumī, *Tetrahedron Lett*, 22, 1417 (1981)
- 5 R H Reuss and A Hassner, *J Org Chem*, 39, 1785 (1974)
- 6 G Greenwood and H M R Hoffmann, *J Org Chem*, 37, 611 (1972)
7. The starting PGI<sub>2</sub> derivatives 1a and 1b were prepared according to the procedure of Johnson et al (see ref 12), and used without purification in the presence of trace amount of triethylamine
- 8 Allyl chloride 2a was also obtained in 55% yield by treatment of 1a with *t*-butylhypochlorite (1.25 equiv, CH<sub>2</sub>Cl<sub>2</sub>, -70°C, 30 min)
- 9 The isomer ratio was estimated by the relative intensity of their <sup>13</sup>C-NMR signals and their C-5 configuration was tentatively determined by the mechanistic consideration
- 10 The crowded concave of this bicyclo[3.3.0] system has been shown by the facile formation of 6(9α), 6(11α)-dioxido-15S-hydroxyprost-13E-enoic acid methyl ester from PGI<sub>2</sub> methyl ester, see K Shimoji, Y Konishi, Y Arai, M Hayashi, and H Yamamoto, *J Am Chem Soc*, 100, 2547 (1978), J Ueda, T Yanagisawa, M Shibasaki, and S Ikegami, *Tetrahedron Lett*, 1978, 2511
- 11 The C-7 configuration was assigned to *S* since the C-7 proton of 5 and 6 appeared as a singlet at 4.79 ppm and 4.69 ppm, respectively, we have observed the coupling constant between 7α- and 8β-proton was less than 1 Hz (see ref 4). The Δ<sup>5</sup>-olefin geometry of 5 and 6 was tentatively assigned to *E* and *Z*, respectively, based on the difference of the C-7 proton chemical shift caused by the deshielding effect of C-5 chlorine atom (see ref 13)
- 12 R A Johnson, F H Lincoln, E G Nidy, W P Schneider, J L Thompson, and U Axen, *J Am Chem Soc*, 100, 7690 (1978)
- 13 S J Rhoads, J K Chattopadhyay, and E E-Waali, *J Org Chem*, 35, 3352 (1970), H O House and V Kramer, *ibid*, 28, 3362 (1963), L M Jackman, "Application of Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry", 2nd ed, Pergamon Press, Oxford, 1969
- 14 The half life was measured by monitoring the intensity of their UV absorption (λ=210 nm)
- 15 M J Cho and M A Allen, *Prostaglandins*, 15, 943 (1978)

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